

# Standardization efforts for Alzheimer CSF biomarkers, and Plasma neurofilament light (NFL) as a biomarker for AD

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# The AD core CSF biomarkers reflect key pathogenic events and are highly clinically validated



AlzBiomarker database - Version 2.0    April 26, 2017

## CSF T-tau

→ Intensity of neurodegeneration

- 188 studies
- 20.600 AD patients and controls
- Effect size 2.48

## CSF A $\beta$ 42

→ Brain amyloid deposition

- 168 studies
- 19.600 AD patients and controls
- Effect size 0.56

## CSF P-tau

→ Phosphorylation state of tau  
and tau pathology

- 116 studies
- 14.300 AD patients and controls
- Effect size 1.88

# Background for starting the Alzheimer's Association CSF program

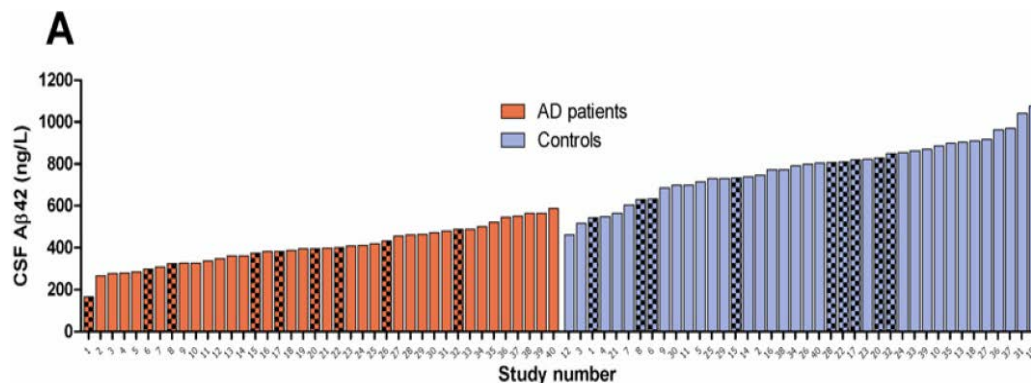


Alzheimer's & Dementia 7 (2011) 386–395

Alzheimer's  
&  
Dementia

## The Alzheimer's Association external quality control program for cerebrospinal fluid biomarkers

Niklas Mattsson<sup>a,\*</sup>, Ulf Andreasson<sup>a</sup>, Staffan Persson<sup>a</sup>, Hiroyuki Arai<sup>b</sup>, Sat Dev Batish<sup>c</sup>, Sergio Bernardini<sup>d</sup>, Luisella Bocchio-Chiavetto<sup>e</sup>, Marinus A. Blankenstein<sup>f</sup>, Maria C. Carrillo<sup>g</sup>, Sonia Chalbot<sup>h</sup>, Els Court<sup>i</sup>, Davide Chiasserini<sup>j</sup>, Neal Cutler<sup>k</sup>, Gunilla Dahlfors<sup>l</sup>, Stefan Duller<sup>m</sup>, Anne M. Fagan<sup>n</sup>, Orestes Forlenza<sup>o</sup>, Giovanni B. Frisoni<sup>p</sup>, Douglas Galasko<sup>q</sup>, Daniela Galimberti<sup>r</sup>, Harald Hampel<sup>s</sup>, Aase Handberg<sup>t</sup>, Michael T. Heneka<sup>u</sup>, Adrianna Z. Henskovič<sup>v</sup>, Sanna-Kaisa Herukka<sup>w</sup>, David M. Holtzman<sup>x</sup>, Christian Humpel<sup>y</sup>, Bradley T. Hyman<sup>z</sup>, Khalid Iqbal<sup>aa</sup>, Mathias Jucker<sup>ab</sup>, Stephan A. Kaeser<sup>ac</sup>, Elmar Kaiser<sup>ad</sup>, Elisabeth Kapaki<sup>ae</sup>, Daniel Kidd<sup>af</sup>, Peter Klivenyi<sup>ag</sup>, Cindy S. Knudsen<sup>ah</sup>, Markus P. Kummer<sup>ai</sup>, James Lui<sup>aj</sup>, Albert Llado<sup>ak</sup>, Piotr Lewczuk<sup>al</sup>, Qiao-Xin Li<sup>am</sup>, Ralph Martins<sup>an</sup>, Colin Masters<sup>ao</sup>, John McAuliffe<sup>ap</sup>, Marc Mercken<sup>aq</sup>, Abhay Moghekar<sup>ar</sup>, José Luis Molinuevo<sup>as</sup>, Thomas J. Montine<sup>at</sup>, William Nowatzke<sup>au</sup>, Richard O'Brien<sup>av</sup>, Markus Otto<sup>aw</sup>, George P. Paraskevas<sup>ax</sup>, Lucilla Parnetti<sup>ay</sup>, Ronald C. Petersen<sup>az</sup>, David Prvulović<sup>ba</sup>, Herman P. M. de Reus<sup>bb</sup>, Robert A. Rissman<sup>bc</sup>, Elio Scarpini<sup>bd</sup>, Alessandro Stefanini<sup>be</sup>, Hilka Soininen<sup>bf</sup>, Johannes Schröder<sup>bg</sup>, Leslie M. Shaw<sup>bh</sup>, Anders Skinningsrud<sup>bi</sup>, Brith Skrogstad<sup>bj</sup>, Annette Spreer<sup>bk</sup>, Leda Talib<sup>bl</sup>, Charlotte Teunissen<sup>bm</sup>, John Q. Trojanowski<sup>bn</sup>, Hayrettin Tumanli<sup>bo</sup>, Robert M. Umek<sup>bp</sup>, Bianca Van Broeck<sup>bq</sup>, Hugo Vanderstichele<sup>br</sup>, Laszlo Vecsei<sup>bs</sup>, Marcel M. Verbeek<sup>bt</sup>, Manfred Windisch<sup>bu</sup>, Jing Zhang<sup>bv</sup>, Henrik Zetterberg<sup>bw</sup>, Kaj Blennow<sup>bx</sup>



→ Large variability for CSF Aβ42 across laboratories

### Variability due to:

Pre-analytical factors	e.g. type of test tube, CSF transfer, freeze-thaw effects,
Analytical factors	e.g. analytical procedures, technician training, run acceptance
Assay manufacturing	e.g. reagent purity, plate coating variability, calibrator stability, lot-to-lot consistency (batch bridging procedure)



# The Alzheimer's Association QC program for CSF biomarkers

- Ongoing project since 2009
- Led by Gothenburg University, funded by the Alzheimer's Association (private sponsor)

## Principle for the QC program:

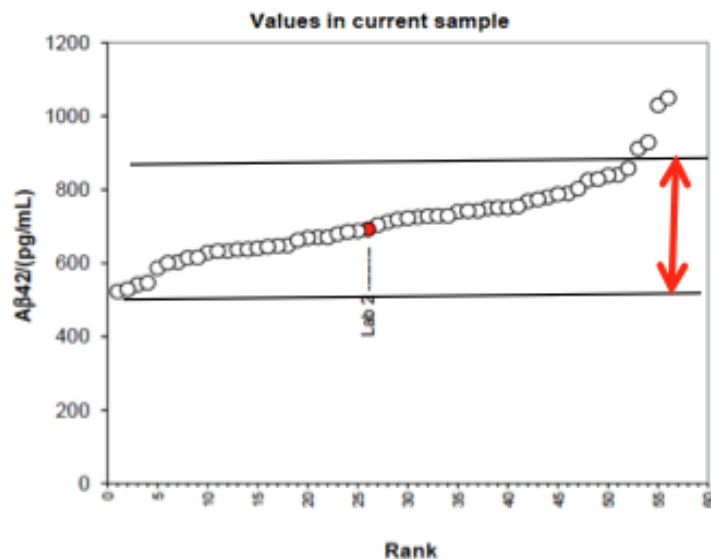
For each round, 3 QC samples (pooled CSF) are sent out  
2 unique samples - for comparisons between labs  
1 identical sample - for comparisons over time

Frequency: 3 times per year



> 90 labs

Gothenburg (Lab 2)		All 56 labs in this round	
Round:	2013:12A	Mean:	717 pg/mL
Result:	693 pg/mL	SD:	110 pg/mL
Method:	INNOTEST	CV:	15,3%

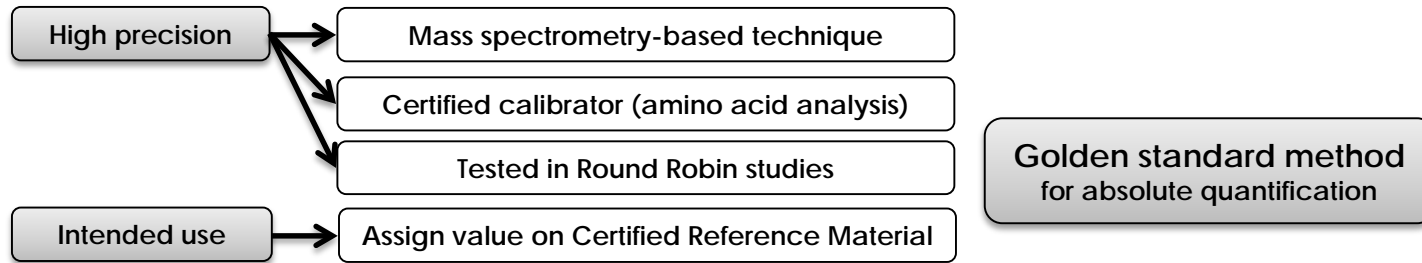


→ Need of standardization efforts

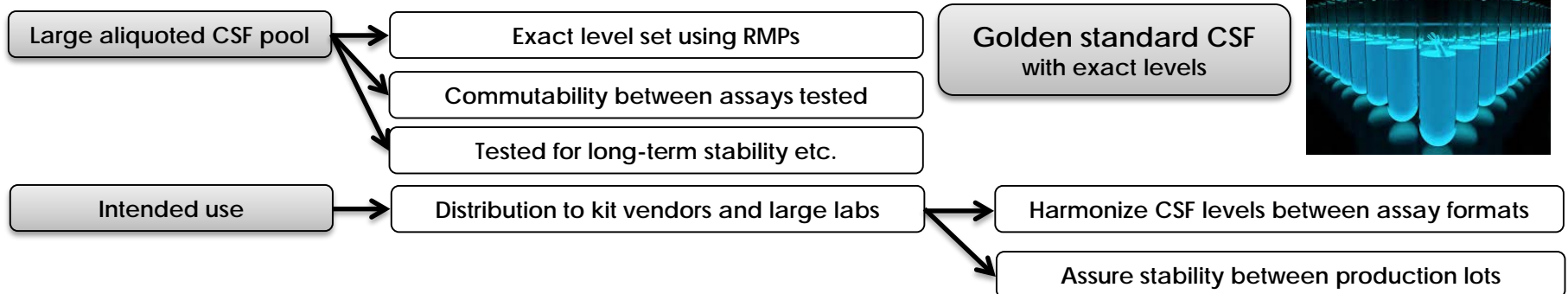


## IFCC Work Group for CSF proteins (IFCC WG-CSF) and Global Biomarker Standardization Consortium (GBSC)

### Reference Measurement Procedure (RMP)

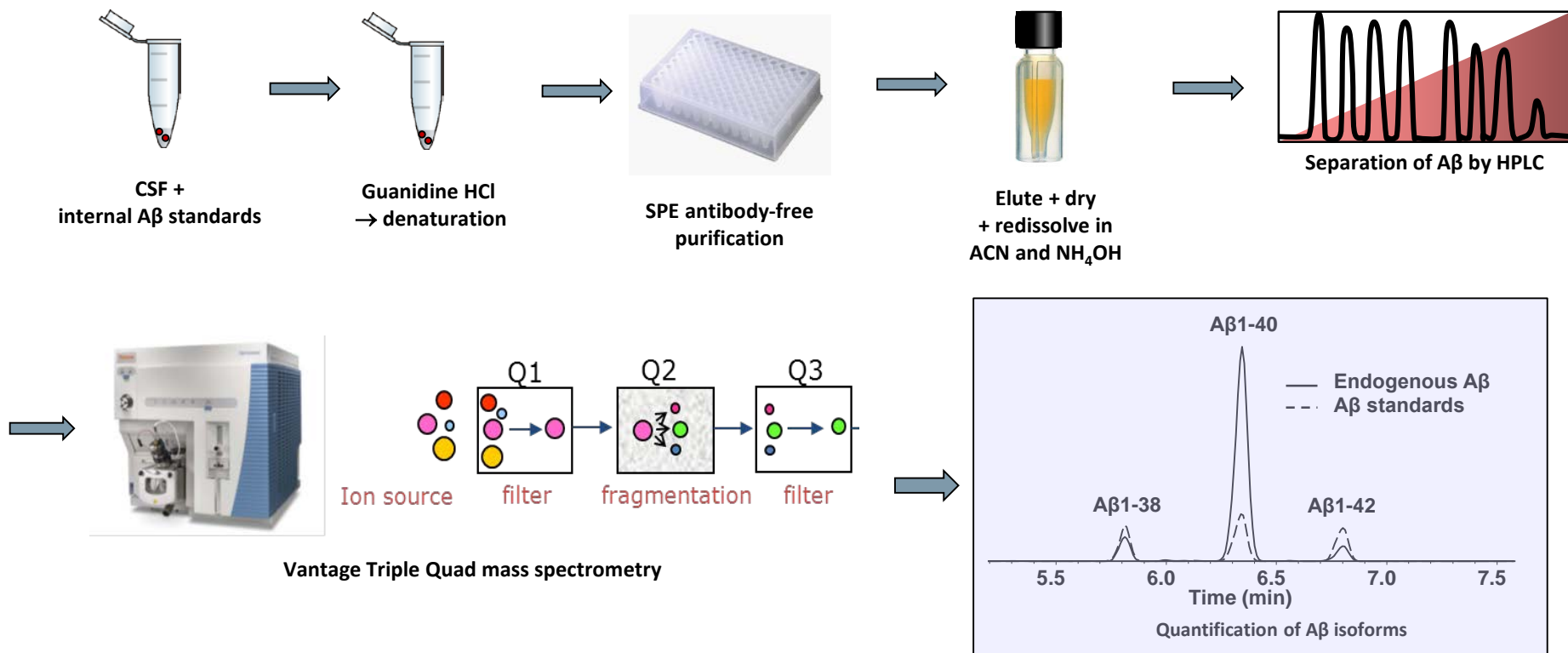


### Certified Reference Material (CRM)



# Reference method for CSF A $\beta$ 42 - Validated "Golden standard" method

- Antibody-free Single Reaction Monitoring (SRM) Triple Quad mass spec method for CSF A $\beta$  isoforms



- Isotope labelled A $\beta$  calibrator added to the CSF sample (and thus processed identically)
- No antibodies involved
- ➔ absolute quantification without interference (matrix effects)

# Mass spectrometry Reference measurement procedure (RMP) for CSF A $\beta$ 42

Clinical Chemistry 60:7  
987–994 (2014)

Proteomics and Protein Markers

## Mass Spectrometry–Based Candidate Reference Measurement Procedure for Quantification of Amyloid- $\beta$ in Cerebrospinal Fluid

Andreas Leinenbach,<sup>1†</sup> Josef Pannee,<sup>2†</sup> Thomas Düllfer,<sup>1</sup> Andreas Huber,<sup>1</sup> Tobias Bittner,<sup>1</sup> Ulf Andreasson,<sup>2</sup> Johan Gobom,<sup>2</sup> Henrik Zetterberg,<sup>2,3</sup> Uwe Kobold,<sup>1</sup> Erik Portelius,<sup>2</sup> and Kaj Blennow<sup>2\*</sup> on behalf of the IFCC Scientific Division Working Group on CSF proteins

Journal of Alzheimer's Disease 41 (2014) 441–451  
DOI 10.3233/JAD-132489  
IOS Press

441

## Qualification of a Surrogate Matrix-Based Absolute Quantification Method for Amyloid- $\beta$ <sub>42</sub> in Human Cerebrospinal Fluid Using 2D UPLC-Tandem Mass Spectrometry

Magdalena Korecka<sup>a</sup>, Teresa Waligorska<sup>a</sup>, Michal Figurski<sup>a</sup>, Jon B. Toledo<sup>a,d</sup>, Steven E. Arnold<sup>b,c</sup>, Murray Grossman<sup>c</sup>, John Q. Trojanowski<sup>a,d</sup> and Leslie M. Shaw<sup>a,d,\*</sup>

## Joint Committee for Traceability in Laboratory Medicine (JCTLM) approvals

Bureau  
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Poids et  
Mesures

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measurement methods/procedures and services

JCTLM  
JCTLM Database  
Laboratory medicine and in vitro diagnostics

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### Result of the search: list of reference measurement methods/procedures

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Your search criteria: Reference measurement methods/procedures; Analyte: amyloid ;  
Analyte category: -; Matrix category: -

Save as PDF file

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#### Results of the search


















##### Isotope dilution mass spectrometry methods for amyloid beta 1-42 in other

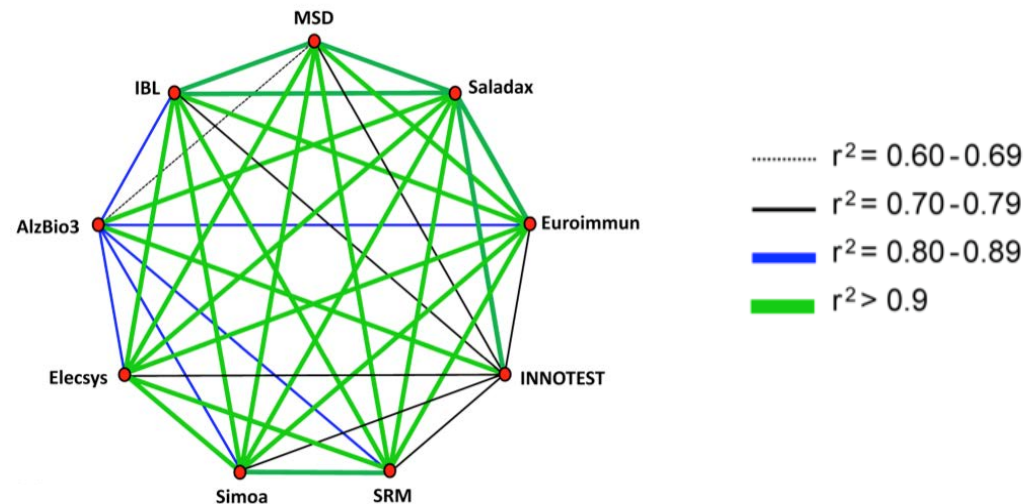
2D-UPLC-tandem mass spectrometric method for analysis of amyloid beta 1-42 in human CSF	
Applicable matrix(es)	frozen human cerebrospinal fluid (CSF)
Full description of technique(s)	Liquid chromatography tandem mass spectrometry, solid phase extraction
Quantity	Mass concentration
Applicable range	100 pg/mL to 3000 pg/mL
Expected uncertainty (level of confidence 95%)	14.3 pg/mL to 355.2 pg/mL
Reference(s)	Qualification of a surrogate matrix-based absolute quantification method for Amyloid $\beta_{42}$ in human cerebrospinal fluid using 2D UPLC-Tandem Mass Spectrometry, Korecka M et al., <i>Journal of Alzheimer's Disease (JAD)</i> , 2014, <b>41</b> (2), 441-451
Comparability assessment study(ies)	Clinical comparison with immunoassay as cited in: Korecka M et al., <i>JAD</i> , 2014, <b>41</b> (2), 441-451 Round robin test on quantification of amyloid- $\beta$ -1-42 in cerebrospinal fluid by mass spectrometry, Pannee J et al., <i>Alzheimer's and Dementia</i> , 2016, <b>12</b> (1), 55-59
Comment(s)	The reference measurement method, C12RMP1, for quantification of A $\beta$ 42 in cerebrospinal fluid was developed and validated by the Biomarker Research Laboratory of Perelman School of Medicine, University of Pennsylvania
JCTLM DB identification number	C12RMP1
* Mass spectrometry-based candidate reference measurement procedure for quantification of A $\beta$ 42 in cerebrospinal fluid	
Applicable matrix(es)	human cerebrospinal fluid
Full description of technique(s)	Isotope dilution mass spectrometry
Quantity	Mass concentration
Applicable range	150 pg/ml to 4000 pg/ml
Expected uncertainty (level of confidence 95%)	15.7 %
Reference(s)	Mass spectrometry-based candidate reference measurement procedure for quantification of A $\beta$ 42 in cerebrospinal fluid, A. Leinenbach et al. on behalf of the IFCC Scientific Division Working Group on CSF proteins (WG-CSF) <i>Clin. Chem.</i> , 2014, <b>60</b> (7), 987-994
Comparability assessment study(ies)	See reference cited above for comparability assessment study
Comment(s)	The reference measurement procedure, C11 RMP9, for quantification of A $\beta$ 42 in cerebrospinal fluid was developed and validated by Roche Diagnostics GmbH in collaboration with the University of Gothenburg
JCTLM DB identification number	C11RMP9

➔ Certified methods for harmonization of results between assays and laboratories

Maria Bjerke, Ulf Andreasson, Julia Kuhlmann, Erik Portelius, Josef Pannee, Piotr Lewczuk, Robert M. Umek, Eugene Vanmechelen, Hugo Vanderstichele, Erik Stoops, Jennifer Lewis, Manu Vandijck, Vesna Kostanjevecki, Andreas Jeromin, Salvatore J. Salamone, Oliver Schmidt, Anja Matzen, Kairat Madin, Udo Eichenlaub, Tobias Bittner, Leslie M. Shaw, Ingrid Zegers, Henrik Zetterberg and Kaj Blennow\*

## Assessing the commutability of reference material formats for the harmonization of amyloid beta measurements

No.	Symbols	Non individual samples	Spiked A $\beta$ 42 concentration, ng/L
		Individual CSF samples	0
1		CSF pool low A $\beta$ 42	0
2		CSF pool high A $\beta$ 42	0
3		aCSF	1000
4		PBS	1000
5		CSF pool low A $\beta$ 42	2000
6		CSF pool low A $\beta$ 42	1000
7		CSF pool low A $\beta$ 42	500
8		CSF pool low A $\beta$ 42	250
9		CSF pool low A $\beta$ 42+0.05% Tween	0
10		CSF pool high A $\beta$ 42+0.05% Tween	0
11		aCSF + 0.05% Tween	1000
12		PBS + 0.05% Tween	1000
13		CSF pool low A $\beta$ 42+0.05% Tween	2000
14		CSF pool low A $\beta$ 42+0.05% Tween	1000
15		CSF pool low A $\beta$ 42+0.05% Tween	500
16		CSF pool low A $\beta$ 42+0.05% Tween	250



- Native CSF pools commutable for almost all method combinations
- CSF pool with spiked A $\beta$ 42 was only commutable at low levels

➔ Three different levels of native CSF pools will be used for three CRMs



# Certified Reference Materials for CSF A $\beta$ 42



**The certification of Amyloid  $\beta_{1-42}$  in CSF in  
ERM<sup>®</sup>-DA480/IFCC, ERM<sup>®</sup>-DA481/IFCC and  
ERM<sup>®</sup>-DA482/IFCC**

Julia Kuhlmann<sup>1</sup>, Sébastien Boulo<sup>1</sup>, Ulf Andreasson<sup>2</sup>, Maria Bjerke<sup>2</sup>,  
Josef Pannee<sup>2</sup>, Jean Charoud-Got<sup>1</sup>, Guy Auclair<sup>1</sup>, Stéphane Mazoua<sup>1</sup>,  
Stefanie Trapmann<sup>1</sup>, Heinz Schimmel<sup>1</sup>, Hendrik Emons<sup>1</sup>, Doris Florian<sup>1</sup>,  
Milena Quaglia<sup>3</sup>, Erik Portelius<sup>2</sup>, Magdalena Korecka<sup>4</sup>, Leslie M. Shaw<sup>4</sup>,  
Mary Lame<sup>5</sup>, Erin Chambers<sup>5</sup>, Hugo Vanderstichele<sup>6</sup>, Erik Stoops<sup>6</sup>,  
Andreas Leinenbach<sup>7</sup>, Tobias Bittner<sup>7</sup>, Rand G. Jenkins<sup>8</sup>, Vesna Kostanjavecki<sup>9</sup>,  
Piotr Lewczuk<sup>10</sup>, Henrik Zetterberg<sup>2</sup>, Ingrid Zegers<sup>1</sup>, Kaj Blennow<sup>2</sup>

Amyloid $\beta_{1-42}$ peptide in human CSF <sup>1)</sup>	Mass concentration	
	Certified value <sup>2)</sup> [ $\mu$ g/L]	Uncertainty <sup>3)</sup> [ $\mu$ g/L]
ERM-DA480/IFCC	0.45	0.07
ERM-DA481/IFCC	0.72	0.11
ERM-DA482/IFCC	1.22	0.18

<sup>1)</sup> As obtained by solid phase extraction and subsequent quantification by liquid chromatography with mass spectrometry detection, according to the reference methods (Leinenbach *et al.* Clin. Chem. 60 (2014) 987-94; Korecka *et al.* J. Alzheimers Dis. 41 (2014) 441-451) [5,6].

<sup>2)</sup> Certified values are values that fulfil the highest standards of accuracy and represent the unweighted mean value of the means of 5 accepted sets of data, each set being obtained in a different laboratory. The certified value and its uncertainty are traceable to the International System of Units (SI).

<sup>3)</sup> The uncertainty is the expanded uncertainty of the certified value with a coverage factor  $k = 2$  corresponding to a level of confidence of about 95 % estimated in accordance with ISO/IEC Guide 98-3, Guide to the Expression of Uncertainty in Measurement (GUM:1995), ISO, 2008 [4].



➔ The CRMs (+dilutions) can be run as a standard curve,  
to calibrate the master calibrator for an immunoassay

➔ Harmonize CSF A $\beta$ 42 readouts between assay formats (different immunoassays)

➔ Candidate RMP for CSF A $\beta$ 40 submitted to JCTLM

➔ One of the CRMs will be used also for CSF A $\beta$ 40

➔ Work in ongoing for a CSF tau mass spec RMP and a CRM for tau

# CSF biomarker assays on fully automated clinical analyzers

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- Fully automated – no variations due to differences in laboratory procedures
  - precise: low between-run, between-batch and between-lab variations
- ➔ Give promise of uniform cut-off levels

- Single sample analysis
  - ➔ analyses can be done directly – better service / fast results to the clinician
  - ➔ no need to await enough samples (n= 35 or 70) to fill an ELISA plate

- Assays available or under development on several platforms:

Cobas Elecsys - Roche

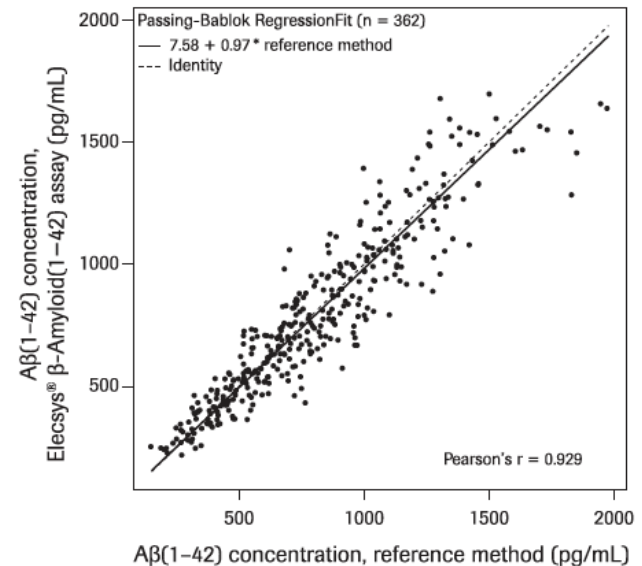
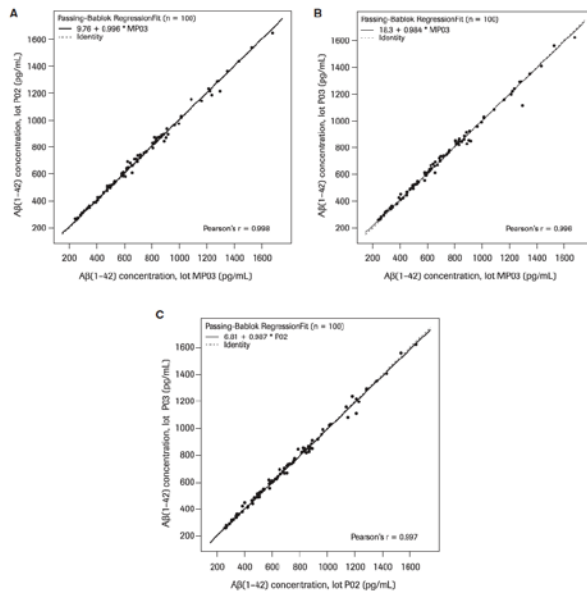
Lumipulse - Fujirebio

RA Analyzer - Euroimmune

## Featured Article

# Technical performance of a novel, fully automated electrochemiluminescence immunoassay for the quantitation of $\beta$ -amyloid (1–42) in human cerebrospinal fluid

Tobias Bittner<sup>a</sup>, Henrik Zetterberg<sup>b,c</sup>, Charlotte E. Teunissen<sup>d</sup>, Richard E. Ostlund, Jr.<sup>e</sup>, Michael Militello<sup>f</sup>, Ulf Andreasson<sup>g</sup>, Isabelle Hubeek<sup>h</sup>, David Gibson<sup>i</sup>, David C. Chu<sup>j</sup>, Udo Eichenlaub<sup>k</sup>, Peter Heiss<sup>l</sup>, Uwe Kobold<sup>m</sup>, Andreas Leinenbach<sup>n</sup>, Kairat Madin<sup>o</sup>, Ekaterina Manuilova<sup>a</sup>, Christina Rabe<sup>a</sup>, Kaj Blennow<sup>b,h</sup>



- ➔ LLOQ 11 pg/mL, linear range 200–1700 pg/mL
- ➔ High lot-to-lot comparability ( $r = 0.995$ )
- ➔ High precision (repeatability CVs of 1.0%–1.6%)

- ➔ standardized to the mass spectrometry RMP for CSF A $\beta$ 42 ( $r = 0.93$ )

- ➔ CSF assays on fully automated analyzers show a marked improvement in performance

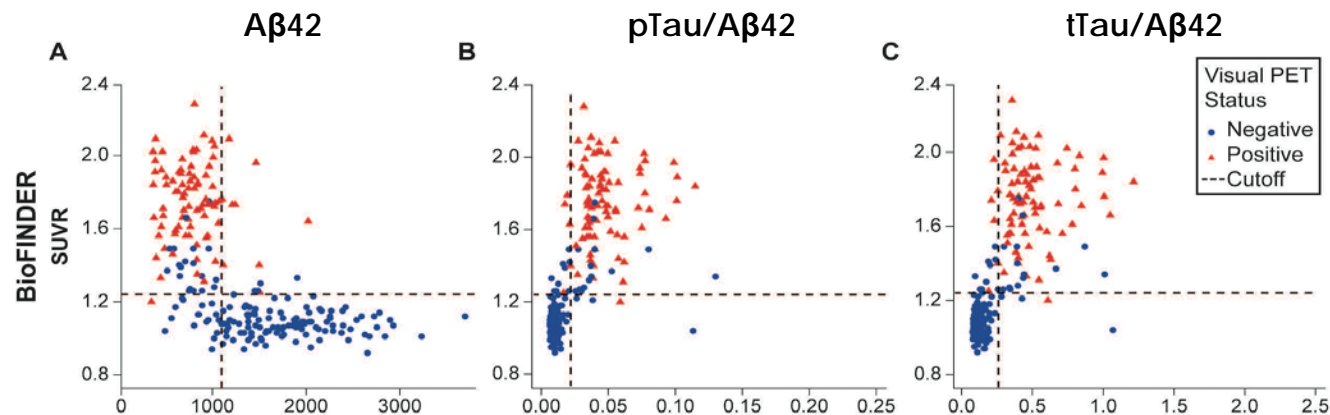
# CSF biomarkers of Alzheimer's disease concord with amyloid- $\beta$ PET and predict clinical progression: A study of fully-automated immunoassays in BioFINDER and ADNI cohorts

Oskar Hansson MD<sup>a,b,\*</sup>, John Seibyl MD<sup>c</sup>, Erik Stomrud MD<sup>a,b</sup>, Henrik Zetterberg MD<sup>d,e,f,g</sup>, John Q. Trojanowski PhD<sup>h</sup>, Tobias Bittner PhD<sup>i</sup>, Valeria Lofke PhD<sup>j</sup>, Veronika Corradini MSc<sup>k</sup>, Udo Eichenlaub PhD<sup>j</sup>, Richard Batrla MD<sup>k</sup>, Katharina Buck PhD<sup>j</sup>, Katharina Zink MSc<sup>j</sup>, Christina Rabe PhD<sup>l</sup>, Kaj Blennow MD<sup>d,e,\*</sup>, Leslie M Shaw PhD<sup>m,\*</sup>, for the Swedish BioFINDER study group<sup>§</sup> and the Alzheimer's Disease Neuroimaging Initiative<sup>||</sup>

## Study design:

Elecsys assays for A $\beta$ 1-42, tTau and Ptau

BioFINDER (n= 277) and ADNI (n= 646)



## Performance of CSF biomarkers vs. visual amyloid PET

Cohort	CSF biomarker	Cutoff	OPA, %
BioFINDER	A $\beta$ (1-42)	1100 pg/mL	79.8 (74.6-84.4)
	pTau/A $\beta$ (1-42)	0.022	89.9 (85.7-93.2)
	tTau/A $\beta$ (1-42)	0.26	89.9 (85.7-93.2)
ADNI	A $\beta$ (1-42)	880 pg/mL	84.4 (81.3-87.1)
	pTau/A $\beta$ (1-42)	0.028	90.3 (87.7-92.4)
	tTau/A $\beta$ (1-42)	0.33	89.2 (86.5-91.5)

Inter-rater PET agreement  
Visual vs. SUVR PET agreement

OPA = 90%  
OPA = 90-91%

→ CSF pTau/A $\beta$ 42 and tTau/A $\beta$ 42 show very high concordance with amyloid PET



# New fully automated techniques in the Alzheimer's Association QC program for CSF biomarkers



Between laboratory CV (percent)				
	INNOTEST® Fujirebio ELISA	Eurolmmune / ADx ELISA	AlzBio3 Fujirebio Luminex	Meso-Scale ECL V-PLEX
Round 14-25 (2014-2018)	β-AMYLOID (1-42)	β-amyloid (1-42)	β-amyloid (1-42)	Human Aβ42
MEAN	16,0	15,2	21,6	17,2
Round 14-25 (2014-2018)	Total tau	Total tau	Total tau	
MEAN	16,0	14,1	15,5	
Round 14-25 (2014-2018)	Phospho tau	Phospho tau		
MEAN	12,3	33,7		

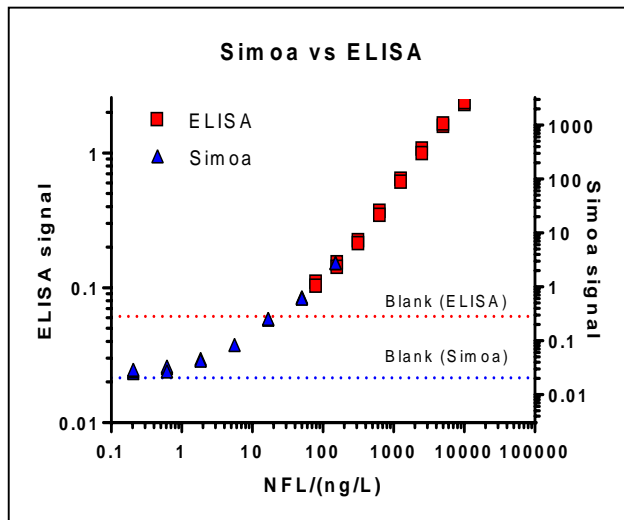
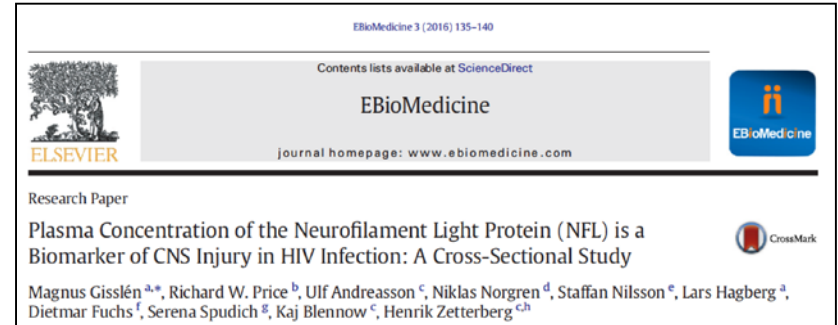
➔ CSF assays on fully automated analyzers show a marked improvement in performance

# Development of the Simoa assay for NFL in blood

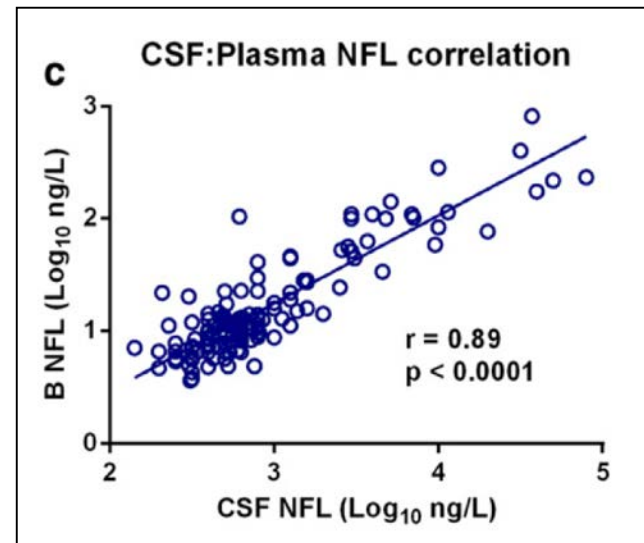
Vinnova (University – Small Enterprise) project in 2013:

Aim to develop a Simoa assay for NFL in blood,  
for use as a biomarker for traumatic brain injury (TBI):

- the same Mabs as in the Uman diagnostics ELISA
- purified bovine NFL as calibrator



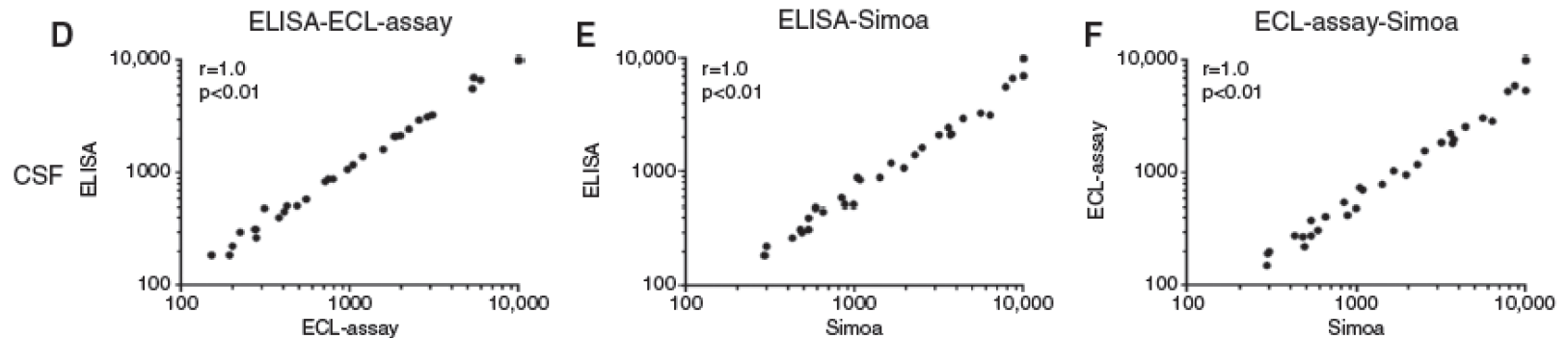
- LLOQ = 0.3 pg/mL (70 pg/mL for ELISA)



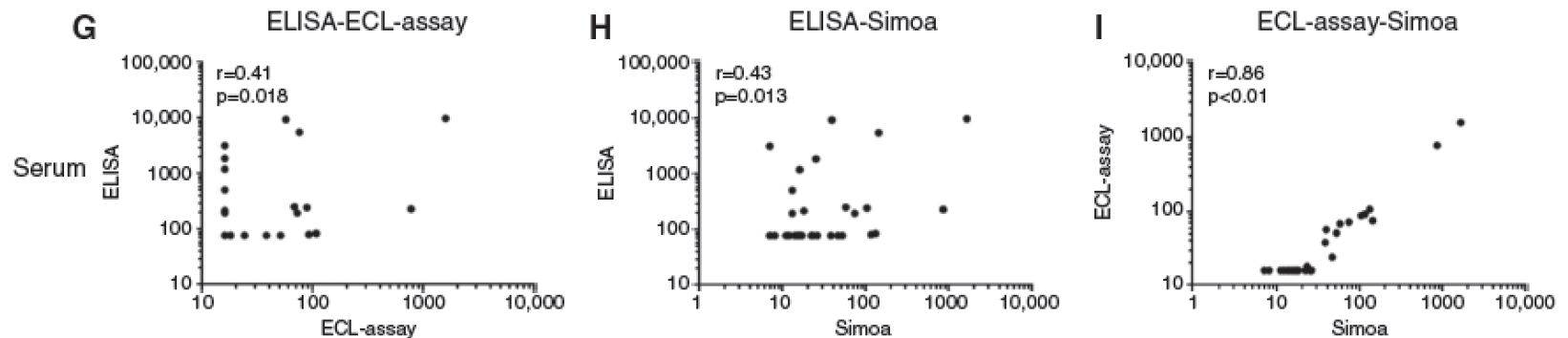
- Very high correlation between plasma and CSF

Jens Kuhle\*, Christian Barro, Ulf Andreasson, Tobias Derfuss, Raija Lindberg, Åsa Sandelius, Victor Liman, Niklas Norgren, Kaj Blennow\* and Henrik Zetterberg\*

## Comparison of three analytical platforms for quantification of the neurofilament light chain in blood samples: ELISA, electrochemiluminescence immunoassay and Simoa



➔ All three assays correlate tightly for CSF samples



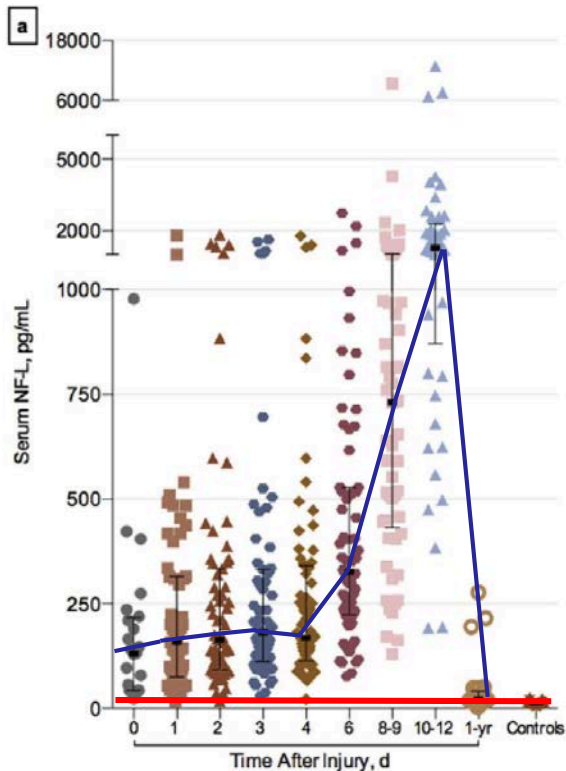
➔ Poor correlations in serum - the ECL and ELISA assay lack analytical sensitivity

OPEN

## Serum neurofilament light protein predicts clinical outcome in traumatic brain injury

Received: 01 July 2016  
Accepted: 17 October 2016  
Published: 07 November 2016

Pashtun Shahim<sup>1</sup>, Magnus Gren<sup>1</sup>, Victor Liman<sup>1</sup>, Ulf Andreasson<sup>1</sup>, Niklas Norgren<sup>2</sup>, Yelverton Tegner<sup>1</sup>, Niklas Mattsson<sup>3</sup>, Niels Andreasen<sup>3</sup>, Martin Öst<sup>6</sup>, Henrik Zetterberg<sup>1,7</sup>, Bengt Nellgård<sup>6</sup> & Kaj Blennow<sup>1</sup>

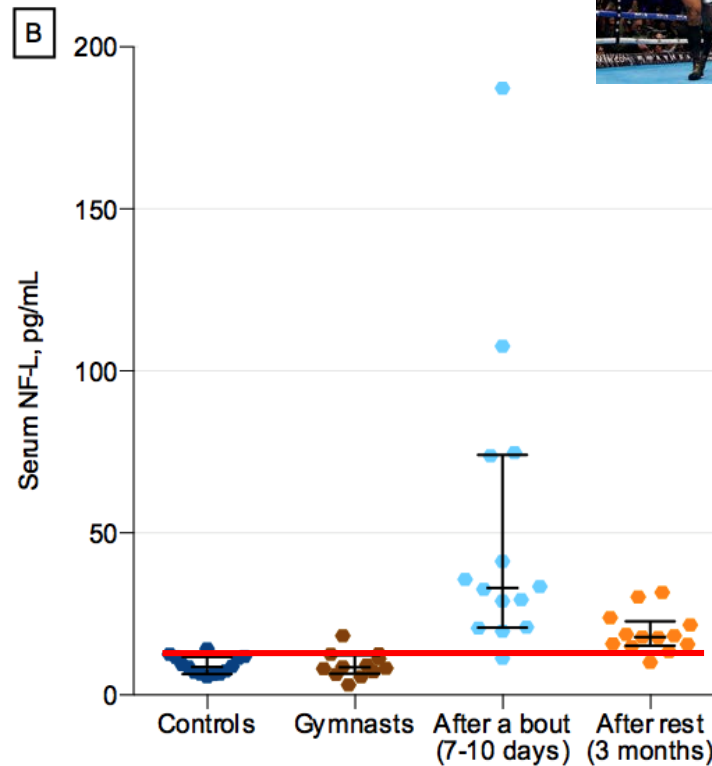


Very marked increase in serum-NFL  
(400 – 4400 % of controls)  
Predicts 1-year clinical outcome

## Serum neurofilament light as a biomarker for mild traumatic brain injury in contact sports

Neurology® 2017;88:1-7

Pashtun Shahim, MD,  
PhD  
Henrik Zetterberg, MD,  
PhD  
Yelverton Tegner, MD,  
PhD  
Kaj Blennow, MD, PhD



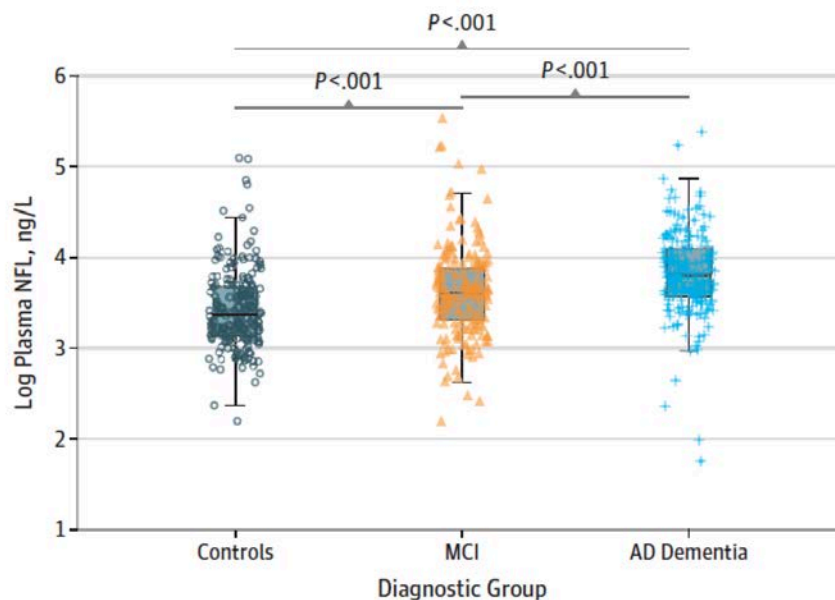
Increased serum NFL after bout  
Higher level with more severe head impact



# Association of Plasma Neurofilament Light With Neurodegeneration in Patients With Alzheimer Disease

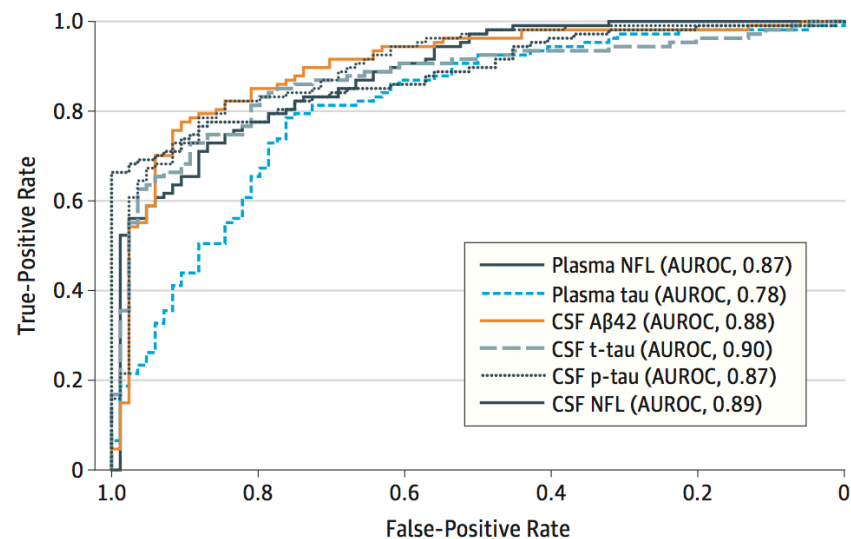
Niklas Mattsson, MD, PhD; Ulf Andreasson, PhD; Henrik Zetterberg, MD, PhD; Kaj Blennow, MD, PhD;  
for the Alzheimer's Disease Neuroimaging Initiative

ADNI cohort: 180 AD dementia, 197 MCI, 193 controls



→ Plasma NFL is increased in AD and MCI

**B** AUROC in AD dementia vs controls

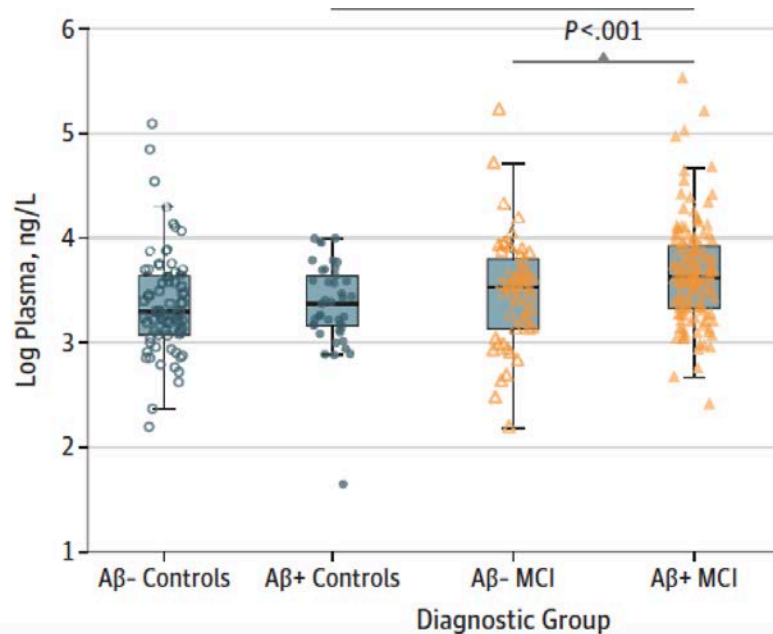


→ High AUC values for AD dementia, similar to CSF A $\beta$ 42, T-tau and P-tau

# Plasma NFL and amyloid load

ADNI cohort

193 controls, 197 MCI



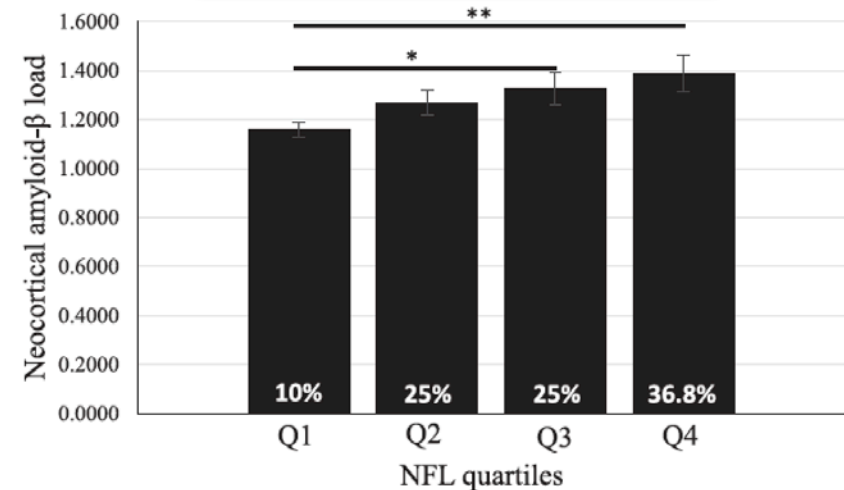
Slightly higher plasma NFL associated with amyloid load in MCI but not in controls

Mattsson N et al. JAMA Neurol 2017

Kerr Anglican Retirement Village Initiative in Ageing Health (KARVIAH) cohort

100 cognitively normal elderly

Within APOE ε4 non-carriers (n=65)



Q1 < 23 Q2 23-34 Q3 34-45 Q4 >45 pg/mL

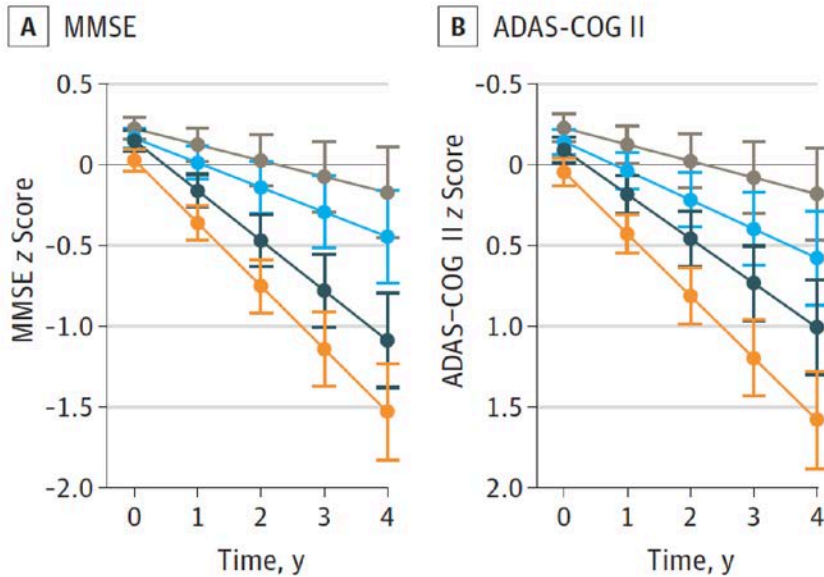
- ➔ No difference in plasma NFL between amyloid positive and negative normal elderly
- ➔ Higher amyloid load in highest plasma NFL quartiles

Chatterjee P et al. J Alzheimer Dis 2018, in press

# Plasma NFL and cognition

ADNI cohort

180 AD dementia, 197 MCI, 193 controls



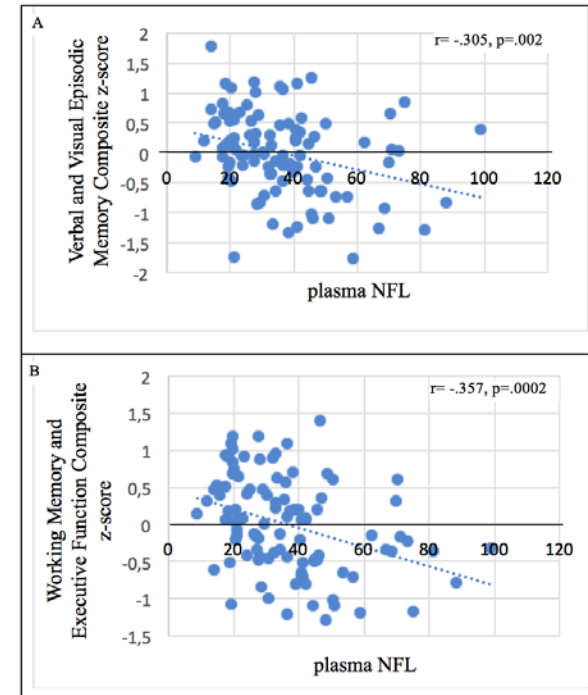
High plasma NFL

associated with baseline MMSE and ADAS-COG  
And predicts future rate of cognitive decline,

Mattsson N et al. JAMA Neurol 2017

Kerr Anglican Retirement Village Initiative  
in Ageing Health (KARVIAH) cohort

100 cognitively normal elderly



High plasma NFL associated with worse cognition  
(episodic memory and executive function)

Chatterjee P et al. J Alzheimer Dis 2018, in press

# Serum neurofilament light in familial Alzheimer disease

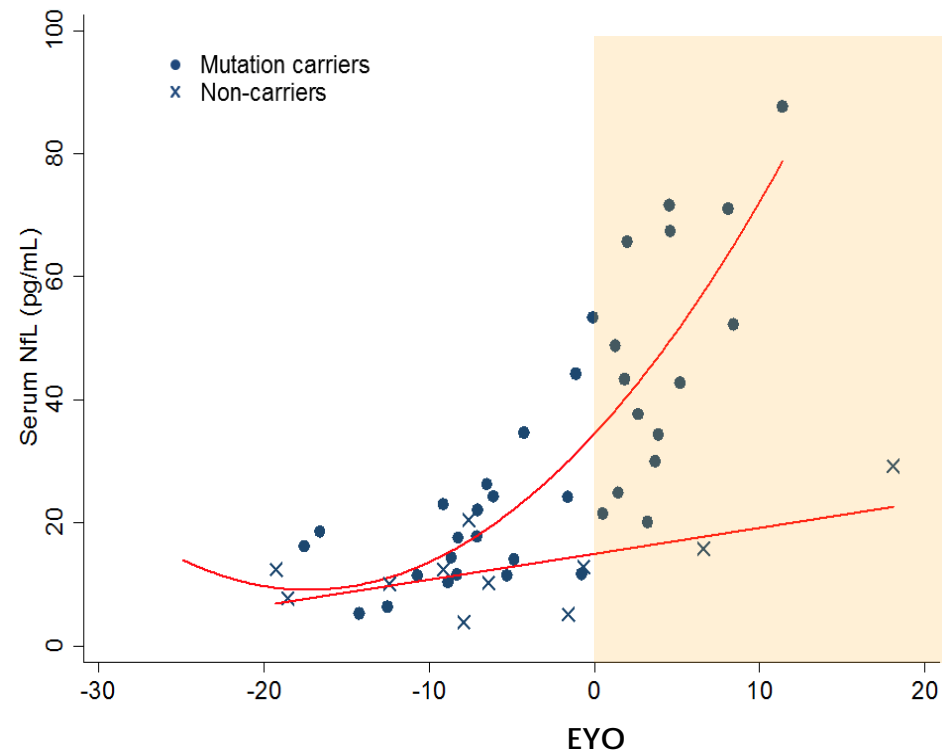
A marker of early neurodegeneration

OPEN

Weston, PSJ, et al *Neurology*® 2017;89:2167-2175

## Study design:

- 18 symptomatic FAD (*APP* or *PSEN*)
- 19 pre-symptomatic carriers
- 11 non-mutation carriers



→ Plasma NFL show promise as a future screening test for neurodegeneration

N.B. Plasma NFL is not AD specific - increase found in several neurodegenerative disorders



# Plasma NFL in other neurodegenerative disorders

## Blood-based NfL

A biomarker for differential diagnosis of parkinsonian disorder

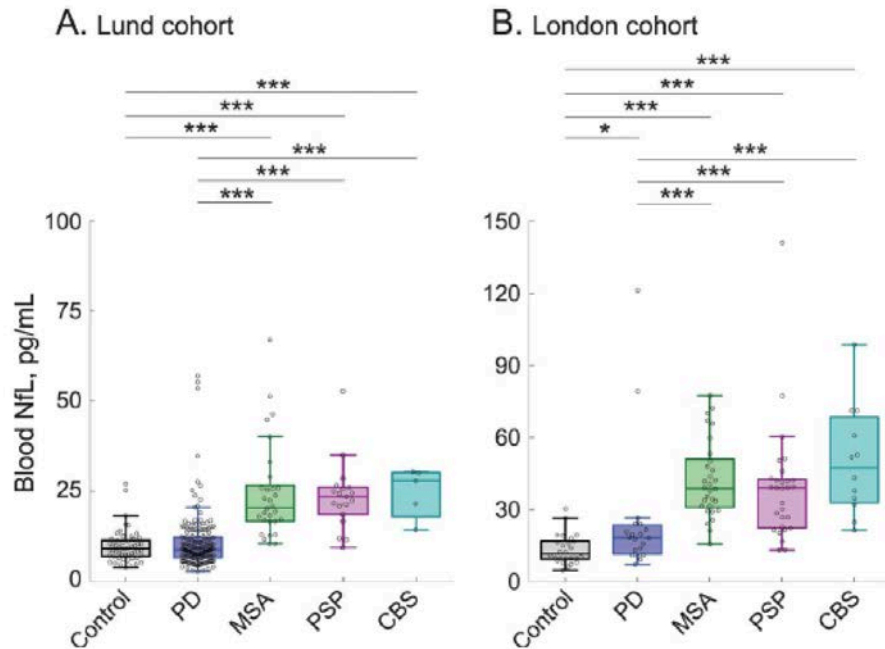
OPEN ▲

*Mattsson N et al, 2017*

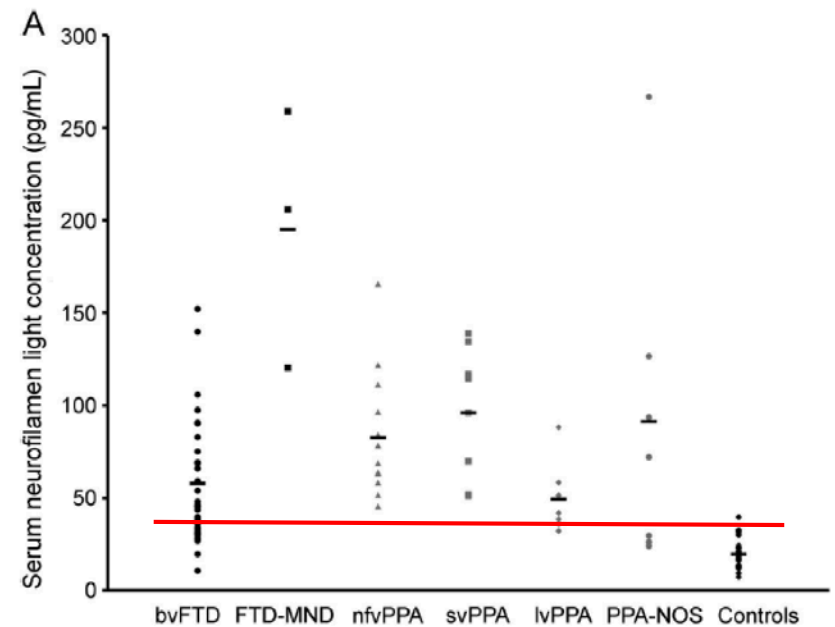
Serum neurofilament light chain protein is a measure of disease intensity in frontotemporal dementia

OPEN

*Rohrer JD et al, 2016*



**Figure 1** Serum neurofilament light chain concentrations in participants by (A) clinical diagnosis and (B) genetic status



→ NFL in blood is a sensitive but disease-unspecific biomarker for neurodegeneration

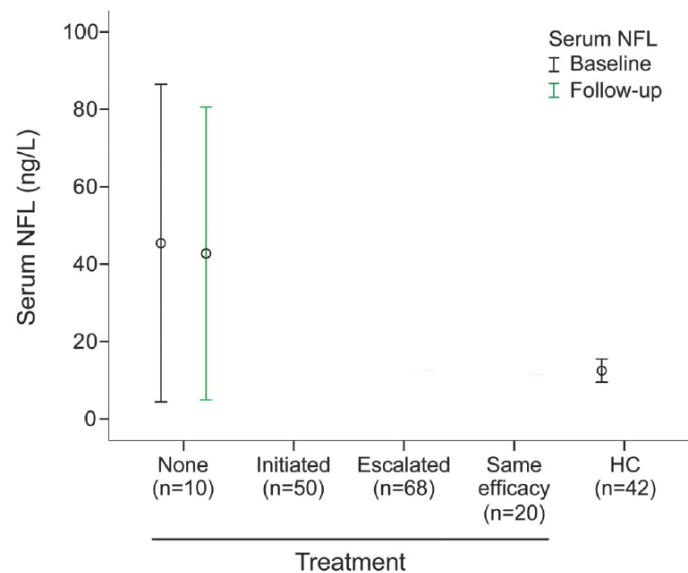
## Monitoring disease activity in multiple sclerosis using serum neurofilament light protein

OPEN

Lenka Novakova *Neurology*® 2017;89:1-8

- 148 MS patients – followed with/without treatment during 12 months
- Less effective DMTs (e.g. interferon- $\beta$ ) more effective DMTs (e.g. fingolimod, natalizumab, or rituximab)

**Figure 1** Serum NFL concentrations in patients with MS at baseline and follow-up and in HCs



→ Reduction in serum NFL with DMTs and with more effective DMTs

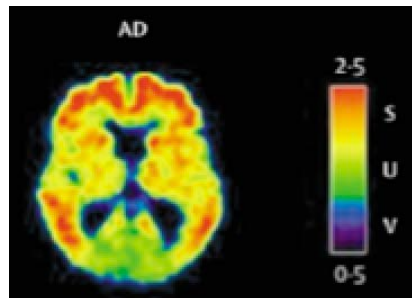
→ Serum NFL may be useful to monitor downstream drug effects on intensity of neurodegeneration

# Blood biomarkers for AD



- Plasma NFL may be useful to screen for neurodegeneration in the first assessment of patients with cognitive symptoms
- May guide clinical management:
  - normal → no further examination
  - increased → admission to specialist clinic

- At the specialist clinic – detailed diagnostic evaluation using 2<sup>nd</sup> grade biomarkers



- Blood NFL may also be valuable in clinical trials
  - to identify and monitor drug effects on neurodegeneration